

AN ANTHRAQUINONE GLYCOSIDE FROM *RHAMNUS PALLASII**

MAKSUT COŞKUN, NEVIN TANKER, AKIYO SAKUSHIMA,† SHIZUKA KITAGAWA† and SANSEI NISHIBE†

Faculty of Pharmacy, University of Ankara, Ankara, Turkey, †Faculty of Pharmaceutical Sciences, Higashi Nippon Gakuen University, Ishikari-Tobetsu, Hokkaido, 061-02, Japan

(Revised received 9 December 1983)

Key Word Index—*Rhamnus pallasii*, Rhamnaceae, α -sorinin, anthraquinone glycoside, physcion-8-*O*- β -primeveroside, 1,8-dihydroxy-3-methyl-6-methoxy-anthraquinone-8-*O*- β -primeveroside

Abstract—A new anthraquinone glycoside, together with α -sorinin, has been isolated from the bark of *Rhamnus pallasii* and its structure elucidated as physcion-8-*O*- β -primeveroside

INTRODUCTION

In a previous paper [1], we reported the isolation of a new dihydroflavonol, pallasin, and seven known flavonoids from the bark of *Rhamnus pallasii* Fisch et May. We now report on the isolation of a known naphthalide, α -sorinin (1), and a new anthraquinone glycoside, physcion-8-*O*- β -primeveroside (2) from the same source.

RESULTS AND DISCUSSION

Compound 1 was identified by direct comparison with α -sorinin isolated from the bark of *R. japonica* Maxim [2]. To the best of our knowledge this is only the second report of the occurrence of α -sorinin in Rhamnaceae.

Compound 2 was recrystallized from methanol to give fine orange needles, mp 258–261°, FDMS m/z 601 ($[M]^+$ ($C_{27}H_{30}O_{14}$) + ^{23}Na). Its IR spectrum showed absorption bands at 3425, 1635, 1600, 1320, 1265, 1220 and 1080 cm^{-1} in potassium bromide, and its UV spectrum showed maximum absorption at 222.5 (4.52), 269 (4.39), 277 (4.38) and 416 (3.94) nm (log ϵ) in methanol, and 266 and 442 nm with addition of sodium hydroxide solution.

Acetylation of 2 with acetic anhydride–pyridine gave the peracetate 3 as pale yellow fine needles (from ethanol), mp 221–223°, $[\alpha]_D^{20}$ –60.5° (CHCl₃). The 1H NMR

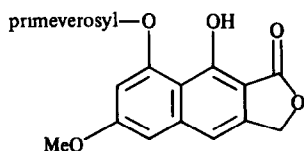
spectrum of 3 showed the presence of six alcoholic acetoxy groups (δ 2.03, 2.08 and 2.10), a phenolic acetoxy group (δ 2.52), an aromatic methoxy group (δ 3.97) and an aromatic methyl group (δ 2.46).

Acid hydrolysis of 2 gave physcion (4), which was identical with an authentic sample. The presence of D-glucose and D-xylose in the hydrolysate was shown by TLC and GC.

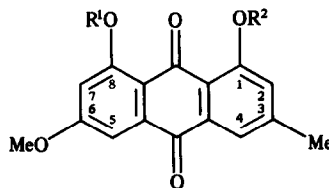
The mass spectrum of 3 contained significant peaks at the m/z values shown in Fig. 1. Those at m/z 547, 259 and 159 suggested the presence of a peracetylxylopyranoglucosyl moiety. The location of the *O*-glycosyl substituent in 2 was established as C-8 by correlation of the chemical shifts of the nuclear protons in the 1H NMR spectrum of 3 with those of physcion-8-*O*- β -glucoside (physcionin) peracetate [3] and physcion-8-*O*- β -gentiobioside peracetate [4] (Table 1).

In the ^{13}C NMR spectrum of 2, the ca 7 ppm downfield chemical shift at C-6 carbon (glc-6) of the glucosyl moiety relative to that of glucose suggested the attachment of the xylosyl moiety at the C-6 carbon of glucose. The 1 \rightarrow 6 linkage in the xylopyrano-glucosyl moiety of 1 had been established by the fact that the physical properties of the disaccharide obtained from 1 by mild hydrolysis and of its phenyllosazone were in good agreement with those of synthetic primeverose (xylopyranosyl-(1 \rightarrow 6)-*O*- β -glucopyranose) [2, 5]. The glycosyl moiety of 2 was shown to be primeverose by correlation of the chemical shifts of its carbon atoms in the ^{13}C NMR spectrum of 2 with those of the primeverosyl moiety of 1 (Table 2).

*Part 2 in the series "Studies on Constituents of *Rhamnus pallasii*". For Part 1, see ref. [1].



1



2 R^1 = primeverosyl, R^2 = H

3 R^1 = (Ac)₆ primeverosyl, R^2 = Ac

4 R^1 = R^2 = H

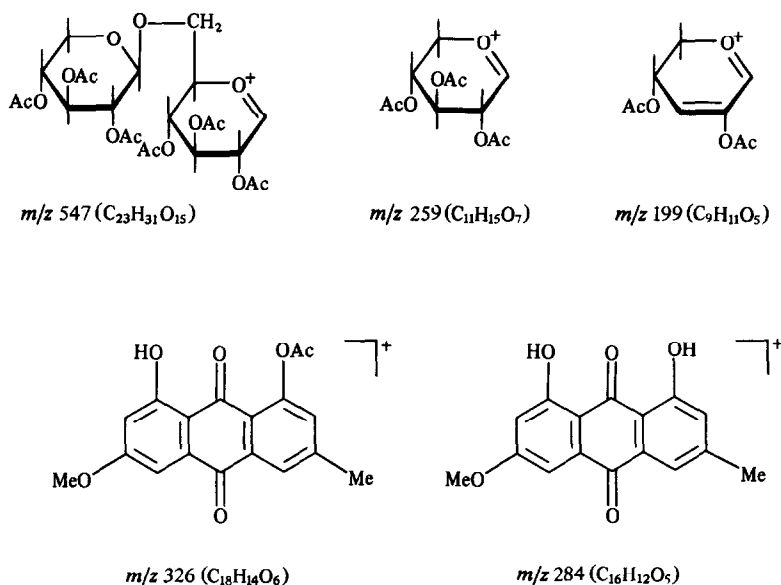


Fig 1

Table 1 Chemical shifts of nuclear protons in 1H NMR spectra of physcion glycoside peracetates

Peracetates	H-2 (s, br)	H-4 (s, br)	H-5 (d, $J = 2.5$ Hz)	H-7 (d, $J = 2.5$ Hz)
Physcion-8- <i>O</i> - β -primeveroside (2)	7.13	7.88	7.45	6.97
Physcion-8- <i>O</i> - β -glucoside*	7.13	7.90	7.43	6.92
Physcion-8- <i>O</i> - β -gentiobioside†	7.19	7.95	7.50	7.00

*Synthesised from physcion and α -acetobromoglucose, mp 164–165°, $[\alpha]_D^{20} -37.2^\circ$ ($CHCl_3$)

†From ref [4]

Table 2 ^{13}C NMR data

Physcion moiety of 2		Primeverosyl moiety		
C		C	2	1
1	164.7	glc-1	100.4	99.8
2	119.2	glc-2	73.2	73.2
3	147.0	glc-3	75.8	76.0
4	124.1	glc-4	69.4	69.7
5	108.3	glc-5	75.8	76.0
6	160.5	glc-6	67.8	67.8
7	107.0	xyl-1	104.0	104.1
8	161.6	xyl-2	73.2	73.2
9	186.3	xyl-3	76.2	76.3
10	181.7	xyl-4	69.4	69.3
11	136.3	xyl-5	65.5	65.5
12	106.9			
13	114.4			
14	131.9			
OMe	56.0			
Me	21.2			

Consequently, the structure of **2** has been established as physcion-8-*O*- β -primeveroside (1,8-dihydroxy-3-methyl-6-methoxy-anthraquinone-8-*O*- β -primeveroside). This is the first report of the occurrence of a physcion diglycoside in Rhamnaceae.

EXPERIMENTAL

1H NMR 90 MHz, $CDCl_3$ with TMS as int. standard, ^{13}C NMR 15 MHz, $DMSO-d_6$, MS direct inlet, 70 eV, ion source temp 200°, GC glass column (3 mm \times 1 m), 1.5% OV-1 on shimalite-W (80–100 mesh), column temp 140–190° (3°/min), injection and detector temp 280°, carrier gas N_2 (20 ml/min).

Plant material *R. pallasii* was collected on Sept. 1980 at Artvin near Ardanuç, Turkey. A voucher specimen is retained in 'Ankara Üniversitesi Eczacılık Fakültesi Herbaryumu' (AEF No 7173).

Isolation Dry powdered bark (100 g) was extracted ($\times 3$) with MeOH. The concentrated extract plus H_2O was extracted successively with Et_2O , $CHCl_3$, $EtOAc$ and BuOH, and the BuOH extract chromatographed on a silica gel column developed with a $CHCl_3$ –MeOH gradient. The fractions were monitored by TLC developed with $CH_3COC_2H_5$ – $EtOAc$ – $HCOOH$ – H_2O – C_6H_6 (4:3:1:1:2, upper layer). The fractions showing a TLC spot

at R_f 0.26 were concentrated to afford crude **2**, whilst those showing a TLC spot at R_f 0.09 were concentrated to afford crude **1**

α -Sorimin (1) Colourless fine needles from EtOH, mp 159–163° IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 3300 (OH), 1720 (chelated CO), 1630, 1610 (arom C=C), 1060, 1030; UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ) 219 (3.55), 250 (4.09) sh, 257 (4.24), 348 (3.36), UV $\lambda_{\text{max}}^{\text{EtOH} + \text{NaOH}}$ nm 241, 264, 365, ^1H NMR ($\text{DMSO}-d_6$) δ 3.84 (3H, s, MeO), 4.16 (1H, d, $J = 7$ Hz, anomeric H), 5.06 (1H, d, $J = 7$ Hz, anomeric H), 5.26 (2H, s, lactone CH_2), 7.00 (1H, d, $J = 3$ Hz, arom H), 7.03 (1H, d, $J = 3$ Hz, arom H), 7.22 (1H, s, arom H)

Acid hydrolysis of physcion-8-O- β -primeveroside (2) Compound **2**, in 10% H_2SO_4 soln, was heated on a water bath for 1 hr then cooled. The mixture was extracted with CHCl_3 . The CHCl_3 layer was washed and evaporated to dryness. The residue was purified by prep TLC (C_6H_6) to give **4**. The aq layer was neutralized with BaCO_3 and the precipitate was filtered off. The filtrate was evaporated to dryness, and the residue was examined by TLC and GC for the presence of D-glucose and D-xylose.

Physcion (4) Dark orange needles from CHCl_3 , mp 207–210° (Found $[\text{M}]^+$ at m/z 284.0678, $\text{C}_{16}\text{H}_{12}\text{O}_5$, requires 284.0683) IR $\nu_{\text{max}}^{\text{KBr}}$ cm^{-1} 1625, 1490, 1370, 1320, 1275, 1220, 1160,

UV $\lambda_{\text{max}}^{\text{EtOH}}$ nm (log ϵ) 223.5 (4.41), 254 (4.14), 264 (4.16), 286 (4.14), 433 (3.99), UV $\lambda_{\text{max}}^{\text{EtOH} + \text{NaOH}}$ nm 212, 229, 255, 304, 503, ^1H NMR (CDCl_3) δ 2.42 (3H, s, Me), 3.87 (3H, s, MeO), 6.57 (1H, d, $J = 2.5$ Hz, H-7), 7.00 (1H, s (br), H-2), 7.26 (1H, d, $J = 2.5$ Hz, H-5), 7.53 (1H, s (br), H-4)

Acknowledgements—We are grateful to Dr H Ina, Tokyo College of Pharmacy, for FD mass spectral data and to Assistant Professor S Yamanouchi, Department of Pharmacy, College of Sciences and Technology, Nihon University, for mass spectral data

REFERENCES

- 1 Sakushima, A, Coşkun, M, Hisada, S and Nishibe, S (1983) *Phytochemistry* **22**, 1677
- 2 Nikuni, Z (1938) *J Agric Chem Soc Japan* **14**, 352
- 3 Steglich, W and Losel, W (1969) *Tetrahedron* **25**, 4391
- 4 Holzschuh, L, Kopp, B and Kubelka, W (1982) *Planta Med* **46**, 159
- 5 Helferich and Rauch (1927) *Ann* **455**, 168